Aquatic Invasive Species Early Detection Monitoring in Lakes

State of Wisconsin Department of Natural Resources

STANDARD OPERATION PROCEDURES

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MONITORING OBJECTIVES

The monitoring objectives are to detect new populations of NR 40 prohibited species and to understand the distribution of NR 40 restricted invasive species within lakes. The Wisconsin Department of Natural Resources (DNR) currently relies on a network of DNR lake biologists and volunteers (i.e. Citizen Lake Monitoring Network) to collect data on the presence and absence of invasive species. The biologist's efforts help evaluate the relationship between water quality and invasive species, and the variable volunteer efforts provide additional reconnaissance, but early detections from these efforts have been incidental. This monitoring will target vulnerable waterbodies for early detection monitoring.

This monitoring strategy may trigger statewide, regional or local implementation of a rapid response strategy, lead to an assessment of abundance and frequency within a waterbody and/or an evaluation of management activities.

Monitoring will be conducted by WDNR staff, partners, and volunteers.

DESIGN

Sample Period: May 15th – September 15th
Frequency: Rotating cycle or as needed

Locations: Boat landings, 5 littoral targeted sites, shoreline, deep hole

Parameters: list of invasive species

Methods: 1. 30 min. search with Dnet & rake (snorkeling optional) at boat landing

2. 10 min. search with Dnet & rake (snorkeling optional) at 5 targeted sites that vary in habitat and include up- and downstream locations and recreation areas

(beaches, resorts, parks, etc.)

2. Shoreline meander

3. Mussel veliger tows (3 at targeted locations)

4. Waterflea Ekman dredges (1 at deep hole)

LAKE SELECTION

The most important factor influencing the introduction and establishment of invasive species are pathways for invasive species to favorable habitats for establishment (Vander Zanden and Olden 2008). The primary vectors for invasive species are anthropogenic, such as public access and recreational opportunity (Leung 2002; Vander Zanden and Olden 2008; Rothlisberger et al. 2011; Latzka personal communication). Disturbed or degraded ecosystems are generally unfavorable for native species and provide niches for invasive species establishment and expansion once a species is introduced (Didham et al. 2005, MacDougall and Turkington 2005, Havel et al. 2005; Johnson et al. 2008).

Our earlier studies suggest that urban land use increase risk of invasive species establishment along streams. Other research suggests that invasive species dispersal is about 15 miles, though more research These protocols will undergo internal DNR review in fall/winter 2021 & external review in winter/spring 2022. If you have major comments, please contact maureen.ferry@wisconsin.gov.

is needed. Therefore, identifying pioneer populations of invasive species in these locations (i.e. urban land use and within 15-mile buffers of know populations) is important to detect new populations and enable response efforts.

Central Office will generate a list of lakes within 15-mile proximity to prohibited species for regional staff to choose from. Biologists will review a list of lakes each winter to monitor the following year. The priority list will be based on the following criteria (in descending order):

- Whether a public boat landing is present;
- ≤15 mi proximity to known occurrences of prohibited or uncommon restricted species; and
- \geq 13% development.

FIELD PREPARATION

Review the AIS Monitoring Field Preparation document which outlines procedures for communication, crews, general safety guidelines, snorkeling guidelines, and ethanol guidelines. https://dnrx.wisconsin.gov/swims/downloadDocument.do?id=239109781

TARGET SPECIES

Table 1. Species targeted for aquatic invasive species surveys. Species with (*) are prohibited early detection species.

Common Name	Latin Name	Species Code
Aquatic Plants		
European Frog-bit*	Hydrocharis morus-ranae	EFB
Yellow Floating Heart*	Nymphoides peltata	YFH
Water Chestnut*	Trapa natans	WC
Brazilian Waterweed*	Egeria densa	BWW
Brittle Naiad*	Najas minor	BRN
Hydrilla	Hydrilla verticillata	HYD
Curly-Leaf Pondweed	Potamogeton crispus	CLP
Fanwort*	Camboba caroliniana	FW
Parrot Feather*	Myriophyllum aquaticum	PF
Eurasian Water Milfoil	Myriophyllum spicatum	EWM
Water Hyacinth*	Eichhornia crassipes and E. azurea	WH
Water Lettuce*	Pistia stratiotes	WL
Didymo*	Didymosphenia geminata	DIDY
Wetland Plants		
Flowering Rush	Butomus umbellatus	FR
Tall Manna Grass*	Glyceria maxima	
Phragmites	Phragmites australis	PHRG
Japanese Knotweed	Polygonum cuspidatum	JK
Giant Knotweed*	Polygonum sachalinense	GK
Purple Loosestrife	Lythrum salicaria	PL

Japanese Hop	Humulus japonicus	JH
Yellow Iris	Iris pseudoacorus	YI
Japanese Stilt Grass*	Microstegium vimineum	JSG
Narrow-Leaf Cattail	Typha angustifolia	NLC
Hybrid Cattail	Typha x glauca	HC
Java Waterdropwort	Oenanthe javanica	
Lesser Celandine	Ficaria verna	
Hairy Willowherb	Epilobium hirsutum	
Graceful Cattail	Typha laxmanni	
Southern Cattail	Typha domingensis	
Bohemian Knotweed	Polygonum x bohemicum	
Golden Creeper	Ernodea littoralis	
European Swamp Thistle	Cirsium palustre	
Poison Hemlock	Conium malculatam	
Invertebrates		
Zebra Mussels	Dreissena polymorpha	ZM
Quagga Mussels*	Dreissena bugensis	QM
Asian Clam*	Corbicula fluminea	AC
Faucet Snails*	Bithynia tentaculata	FS
Spiny Waterfleas*	Bythotrephes longimanus	SWF
Chinese Mystery Snails	Cipangopalundina chinesis	CMS
Banded mystery Snails	Viviparus georgianus	BMS
New Zealand Mudsnail	Potamopyrgus antipodarum	NZMS
Red Swamp Crayfish*	Procambarus clarkii	RSC
Rusty Crayfish	Orconectes rusticus	RC

FIELD GUIDES

For guidance on target species identification and habitat preference, please review the attached and the following guides:

- AIS Identification Guide (multi-page) and AIS Identification Guide (one-page)
- <u>Crayfish of Wisconsin Guide (2017)</u> (multi-page)
- Citizen Lake Monitoring Network Aquatic Invasive Species Monitoring Protocol
- Field guide to Wisconsin Streams
- Key to Wisconsin Freshwater Snails

EQUIPMENT

All lakes

- Gazetteer
- Bathymetric map of lake

- Boat
- Boat ladder (for easily exiting and entering the boat in deep water)
- GPS

- Waterproof camera
- Polarized sunglasses
- Waterproof paper (to print monitoring forms and specimen labels on)
- D-net
- Aquatic plant rake on a rope and pole ~ (detailed instructions in <u>Recommended</u> <u>Baseline Monitoring of Aquatic Plants in</u> Wisconsin
- 54 μm plankton net (veliger samples)
- Ekman grab w/ line and messenger (waterflea samples)
- Wash bucket w/ 500 micron filter bottom
- Spray bottle
- Gallon or quart sized zip-locs (1 per Ekman grab)
- Sonde or Hydrolab (to measure conductivity)
- Secchi disk
- Depth finder (in boat or handheld)
- Mesh bag (nylons or fruit mesh bags can be used) – to place specimens in underwater
- Sorting trays for plants and inverts
- Ziploc bags for plant preservation (can be reused)
- Plastic bottles for benthic invertebrate specimens (~3 per lake)
- Bottle labels printed on waterproof paper

Turbid lakes

 Latex or other gloves to protect hands from blue-green algae.

Clear lakes

- Wetsuits
- Dive flag, float and anchor
- Snorkel, mask, fins and weight belt
- Snorkel mask clearing solution
- Stopwatch

Disinfection Equipment

- Long-handled stiff bristled brush
- Portable Jiffy steam cleaner
- Jugs for clean water
- Tubs for equipment disinfection
- Backpack or handheld sprayer
- Bleach solution
- Black plastic bags
- Nitrile gloves
- Emergency eyewash station or eye wash solution
- Eye protection meeting ANSI Z87 (safety sunglasses are acceptable)
- Splash goggles and/or a face shield
- Dust mask respirator (optional requires training)
- Chlorine solution and/or Virkon® Aquatic

Preservation and Vouchering

- Water proof camera
- Ethanol (non-denatured preferred) for veliger sample preservation and prohibited species
- Plant press, newspaper and herbarium paper for prohibited plant species

FIELD METHODS

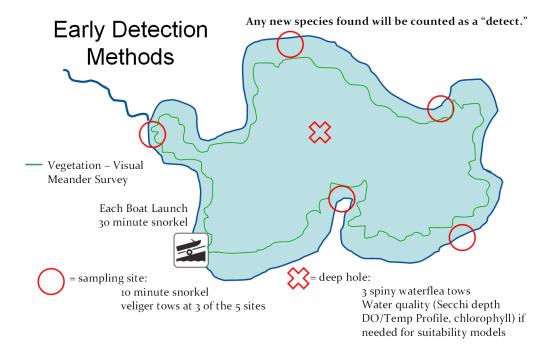


Figure 1 Schematic of AIS Early Detection Lake Survey Design (Latzka and Van Egeren 2010). This map does not accurately illustrate tows and has additional water quality information that is not collected during these surveys

Data Collection

Collect data using appropriate forms on waterproof paper.

- Aquatic Invasive Species Early Detection Monitoring Form
- Mussel Veliger Tow Labslip (generated in SWIMS)
- Water Flea Tow Labslip (generated in SWIMS)

Photo/Specimen Collection Protocol

- Collect photographs of all new AIS occurrences for verification purposes. Follow the DNR
 Photographing Aquatic and Wetland Invasive Species Guidance:
 https://dnrx.wisconsin.gov/swims/downloadDocument.do?id=145712698.
- Collect specimens of <u>prohibited</u> or unusual invasive species for vouchering. Collect about 5 intact specimens that contain all the identifying characteristics. While in the boat, store specimens in separate sample bottles or in Ziploc bags with water or a wet paper towel.
- Upon completion of the survey, specimens should be properly preserved and submitted to the Regional DNR AIS Coordinator as soon as possible.
- Record data on the Early Detection Form.

Boat Landing Search(es)

- Survey all public boat landings (public and commercial). Do not include small, backyard boat ramps. Record the location of each boat landing from the shoreline in decimal degrees using a GPS (datum WGS84) whether or not any AIS are found.
- Each landing is searched for 30 minutes using snorkeling, D-nets, rakes, and surveying shoreline. Snorkel for 15 minutes covering an area of shoreline 200' long out to the maximum depth of plant growth or 100' from shore. For distance reference, baseball bases are 90' apart and a football field is 300' long. Spend 15 minutes using the D-net and rake in the shallow water and also examining the shoreline for 200'.
- For QAQC purposes, <u>collect specimens of all AIS recorded</u>.
- Do not swim in water that looks like "pea soup", green or blue paint, or that has a scum layer or puffy blobs floating on the surface. If there is poor visibility or safety is a concern (e.g. bluegreen algae bloom), do not snorkel. Instead, analyze rake tows and D-net samples for about 30 minutes within the defined area (200' shoreline and 100' from shore).
- Record data on Early Detection form.

Targeted Search Sites

- Stop at each targeted search site and conduct 10-minute snorkel searches. If there are two
 people, one person should snorkel for 5 minutes while the other person examines the shallows
 and shoreline for 5 minutes (while also keeping watch on the snorkeler). If you find additional
 appropriate search sites during the shoreline meander, you can add additional 10-minute
 searches or replace the sites that you pre-selected.
- For QAQC purposes, collect specimens of all AIS recorded.
- If there is poor visibility or safety is a concern (e.g. blue-green algae bloom), do not snorkel. Instead, at each site analyze rake tows and D-net samples for about 10 minutes.
- Record the site number and location (center of site at shoreline in decimal degrees) of each site
 on the datasheet whether or not any AIS are found. Record the name of any species found and
 density rating (as defined on the datasheet). Record data on Early Detection Form.

Meander Survey

- During the meander survey, drive boat slowly between target sites and look for aquatic invasive plants, mussels, and snails in the water and along the shoreline. Meander between shallow water and maximum rooting depth or 100' from shore whichever comes first.
- If not snorkeling the target sites due to poor visibility or safety concerns, visibility will likely be difficult during the meander survey also. To sample during the meander survey in low visibility, collect 50 rake tows/D-net samples during the meander survey.
- Record the site number, location in decimal degrees and the name and density (as defined on the back of the datasheet) of each AIS occurrence observed. However, only collect separate GPS points from discretely different beds or populations of invasive species.
- If three discrete locations of a certain species are found either at targeted search sites and/or during the meander survey stop recording new locations during the meander survey. Three discrete locations of one species will indicate that the species is established in the lake.
- Collect specimens of all AIS reported for QAQC purposes.
- Record data on Early Detection Form.

Waterflea Tows

 See the waterflea collection protocol: https://dnrx.wisconsin.gov/swims/downloadDocument.do?id=234380921

Veliger Tows

 See the mussel veliger collection protocol: https://dnrx.wisconsin.gov/swims/downloadDocument.do?id=234380927.

FOLLOWING FIELD PROTOCOL

Equipment Disinfection

 Review the manual code and best management practices on the <u>Boat, Gear and Equipment</u> <u>Decontamination and Disinfection Website</u>

Verification

 Follow the AIS Early Detection Monitoring Verification and Voucher Guidance for all suspected new AIS occurrences: https://dnrx.wisconsin.gov/swims/downloadDocument.do?id=145713313

Data Entry

 Enter Early Detection form data into SWIMS <u>once each week</u> in the "Aquatic Invasive Species Early Detection Surveys" project.

Contact your Regional AIS Coordinator if you have questions about SWIMS data entry. The
regional DNR AIS Coordinator will contact the Statewide AIS Monitoring Lead if they have
questions.

Notification

 If AIS not previously observed in a waterbody are found, follow the Department's communication protocol:

http://dnr.wi.gov/water/wsSWIMSDocument.ashx?documentSeqNo=118813642.

Voucher

 Follow the AIS Early Detection Monitoring Voucher Guidance for all new NR40 prohibited or unusual species occurrences:

https://dnrx.wisconsin.gov/swims/downloadDocument.do?id=145713415

